

· 论 文 ·

一种查耳酮类化合物的合成方法

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摘 要 建立一种以取代苯乙酮和取代苯甲醛为原料, 采用多聚磷酸/浓硫酸体系催化合成查耳酮类化合物的方法, 并对反应条件进行优化。结果表明, 最优反应条件为多聚磷酸 5 equiv.、浓硫酸 20 equiv., 1,4-二氧六环作溶剂, 90 °C 氮气保护下反应 2 h。在优化的反应条件下, 以较好的得率合成了 12 个查耳酮类化合物。所有化合物结构通过 IR、HRMS、¹H NMR 和 ¹³C NMR 进行表征。

关键词 查耳酮类化合物; 缩合反应; 多聚磷酸/浓硫酸; 苯乙酮; 苯甲醛; 合成

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A method for the synthesis of chalcones

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Abstract In this paper, a new method for catalytic synthesis of chalcones from substituted acetophenone and substituted benzaldehyde in polyphosphoric acid/concentrated sulfuric acid system was proposed, and the reaction conditions were optimized. The results showed that the optimized reaction conditions were determined as polyphosphoric acid of 5 equiv. and concentrated sulfuric acid of 20 equiv., with 1,4-dioxane as solvent at 90 °C for 2 h under nitrogen protection. Twelve chalcones were synthesized with good yield. All target compounds were characterized by IR, HRMS, ¹H NMR and ¹³C NMR.

Key words chalcones; condensation; polyphosphoric acid/H₂SO₄; acetophenones; benzaldehydes; synthesis

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查耳酮是许多天然存在的化合物中最常见的骨架之一, 研究发现查耳酮类化合物的生物活性

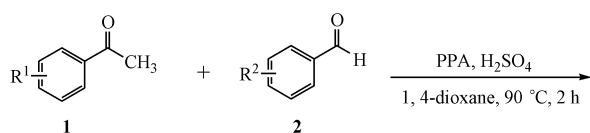
较为广泛, 例如肿瘤、炎症和糖尿病。查耳酮类化合物美托查酮(metochalcone)和索法酮(sofalcone)

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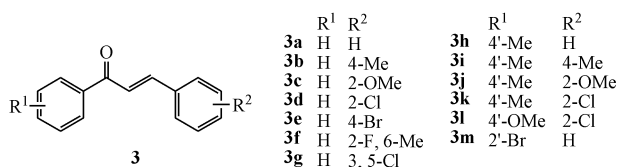
曾分别作为抗胆碱药和抗溃疡药应用于临床。查耳酮化合物由于自身结构的柔性能与诸多受体相结合,从而体现出了许多生物活性,如抗蛲虫、抗过敏、抗肿瘤、抗氧自由基、抗菌、抗病毒、抗溃疡和解痉。合成查耳酮类化合物经典的合成方法为碱或酸催化的 Claisen-Schmidt 缩合^[1]。常使用的碱催化剂有氢氧化钠、氢氧化钾、碱土金属氢氧化物、双(三甲硅基)酰胺锂和哌嗪。酸催化剂有氯化氢、三氯化铝、四氯化钛、氯化钨、三氟化硼、对甲苯磺酸。也有文献报道了研磨法^[2]、微波加热法^[3]以及水作为溶剂相转移催化法^[4]合成查耳酮类衍生物。但是 Claisen-Schmidt 缩合存在反

应时间较长、常伴有副产物或反应不彻底导致难以分离的缺点^[5]。除了经典缩合反应外,交叉偶联法(Suzuki 反应^[5]、Heck 反应^[6]、Julia-Kocienski 反应^[7]、Wittig 反应^[8])和 Friedel-Crafts 酰化反应也常用于查耳酮类化合物的合成^[9],但总体得率不理想。因此,开发简便、高效、反应条件温和的查耳酮类化合物的合成方法具有重要意义。本研究以 1,4-二氧六环(1,4-dioxane)为溶剂,PPA/H₂SO₄体系催化苯乙酮和苯甲醛缩合成系列查耳酮类化合物(路线1)。此方法操作简单、反应条件温和、产率高且所使用的催化剂成本低。



Scheme 1 Synthesis route of chalcones

PPA: Polyphosphoric acid



1 试剂和仪器

多聚磷酸(PPA,纯度大于80%,海业联联合化工有限责任公司);浓硫酸(纯度98%,重庆川江化学试剂厂);1,4-二氧六环[纯度99.7%,重庆川东化工(集团)有限公司];其他试剂均为市售分析纯。

INOVA 600 MHz 核磁共振仪(TMS 内标)、Impact HD Q-TOF 型高分辨质谱仪(德国 Bruker 公司);红外光谱仪(天津港东科技股份有限公司);XT-4 型显微熔点测定仪(北京泰光仪器有限公司)。

2 目标化合物的合成

2.1 反应条件的优化

以苯甲醛(0.8 mmol)和苯乙酮(1.0 mmol)为反应底物,优化反应条件。分别考察 PPA 用量、浓硫酸用量、反应温度、反应时间、酸的种类以及溶剂种类对反应的影响,筛选出最优反应条件。

2.2 反应底物的拓展

以化合物 3a 为例,在优化好的实验条件基础上,取 PPA(5.0 mmol)加入 20 mL 圆底烧瓶中,再依次加入 1,4-二氧六环(3.5 mL)、浓硫酸(20.0

mmol)、苯甲醛(0.8 mmol)和苯乙酮(1.0 mmol),于 90 °C 氮气保护下油浴搅拌加热反应,2 h 反应结束。用乙酸乙酯、饱和食盐水萃取,有机相减压浓缩后经柱色谱分离纯化得到化合物 3a,洗脱剂为石油醚-乙酸乙酯(50:1)。以相同条件合成查耳酮化合物 12 个,所有化合物经核磁共振氢谱(¹H NMR)、核磁共振碳谱(¹³C NMR)、高分辨质谱(HRMS)、红外波谱(IR)和熔点仪表征,理化性质数据列于表 1,核磁表征数据列于表 2。

3 结果与讨论

3.1 反应条件的优化

多聚磷酸(PPA)在有机合成中用作失水剂、环化剂、酰化剂,是缩合、环化、重排、取代等反应的催化剂或溶剂。本研究以苯甲醛(0.8 mmol)和苯乙酮(1.0 mmol)为反应底物,优化反应条件。考察了 PPA 用量、浓硫酸用量、反应温度、反应时间、酸的种类以及溶剂种类对反应的影响。薄层色谱(TLC)监测反应进程,反应结束后分离得到目标化合物,比较了不同条件下的产率,从而筛选出最佳反应条件,结果列于表 3。

PPA 用量为 10 equiv.,浓硫酸用量为 15 equiv.,以二氧六环为溶剂,90 °C 搅拌反应 2 h,以

Table 1 Appearance,yields,melting points,HRMS and IR data of compounds **3a**–**3l**

Compd.	Appearance	Yield/%	mp/°C	HRMS, m/z [M+H] ⁺ (calcd.)	IR(KBr), $\nu_{\max}/\text{cm}^{-1}$
3a	White solid	96	51–54(54–56 ^[10])	208.112 8(208.088 8)	1 648, 1 585, 1 563, 819, 760, 720, 690
3b	Yellow solid	95	92–95(95–96 ^[11])	222.105 6(222.104 4)	1 642, 1 575, 1 555, 810, 765, 728, 693
3c	Yellow solid	90	63–64(58–60 ^[12])	238.146 7(238.099 4)	1 667, 1 560, 1 588, 849, 754, 740, 686
3d	Yellow solid	82	54–55(50–52 ^[13])	242.125 8(242.049 8)	1 682, 1 577, 1 542, 807, 752, 701, 640
3e	Light red solid	83	120–122(122–123 ^[14])	285.864 7(285.999 3)	1 686, 1 592, 1 535, 820, 778, 734, 682
3f	Yellow solid	86	77–79	240.115 8(240.095 0)	1 661, 1 581, 1 569, 849, 780, 742, 710
3g	Yellow solid	80	75–78	276.102 4(276.010 8)	1 650, 1 580, 1 572, 818, 750, 710, 688
3h	Light red solid	91	60–64(50–51 ^[15])	222.148 7(222.104 5)	1 653, 1 574, 1 562, 852, 776, 740, 696
3i	Light red solid	91	124–127(128–130 ^[16])	236.158 8(236.120 1)	1 679, 1 595, 1 576, 855, 774, 738, 712
3j	Red oil	92	/	252.164 1(252.115 0)	1 687, 1 592, 1 525, 841, 777, 741, 682
3k	Light red solid	88	74–76	256.126 6(256.065 4)	1 669, 1 594, 1 542, 852, 760, 740, 668
3l	Orange solid	86	79–81(79–80 ^[17])	272.108 6(272.060 4)	1 622, 1 590, 1 560, 849, 761, 725, 687

Table 2 ¹H NMR and ¹³C NMR data of compounds **3a**–**3l**

Compd.	¹ H NMR(600 MHz,CDCl ₃), δ	¹³ C NMR(150 MHz,CDCl ₃), δ
3a	8.03(s, 1H, Ph-H), 8.02(d, J =1.3 Hz, 1H, Ph-H), 7.82(d, J =15.7 Hz, 1H, β -H), 7.65(dd, J =6.5, 3.0 Hz, 2H, α -H, Ph-H), 7.59(t, J =7.4 Hz, 1H, Ph-H), 7.55(s, 1H, Ph-H), 7.52(dd, J =12.4, 4.4 Hz, 3H, Ph-H), 7.43(dd, J =5.0, 1.8 Hz, 3H, Ph-H)	190.93, 145.21, 138.57, 135.24, 133.14, 130.90, 129.32, 128.98, 128.86, 128.80, 122.46
3b	8.03(s, 1H, Ph-H), 8.01(d, J =1.4 Hz, 1H, Ph-H), 7.80(d, J =15.7 Hz, 1H, β -H), 7.58(d, J =7.4 Hz, 1H, α -H), 7.55(d, J =8.1 Hz, 2H, Ph-H), 7.52–7.50(m, 2H, Ph-H), 7.49(d, J =4.0 Hz, 1H, Ph-H), 7.23(d, J =7.9 Hz, 2H, Ph-H), 2.40(s, 3H, -CH ₃ -H)	191.00, 145.29, 141.43, 138.70, 133.00, 132.50, 130.05, 128.93, 128.81, 121.44, 21.88
3c	8.05(d, J =2.0 Hz, 1H, Ph-H), 8.04(d, J =2.0 Hz, 1H, Ph-H), 7.81(d, J =15.7 Hz, 1H, β -H), 7.64(dd, J =7.4, 2.0 Hz, 2H), 7.55(d, J =15.7 Hz, 1H, α -H), 7.42–7.40(m, 3H, Ph-H), 6.99(d, J =8.9 Hz, 2H, Ph-H), 3.89(s, 3H, -OCH ₃ -H)	189.07, 163.78, 144.31, 135.43, 131.44, 131.16, 130.67, 129.26, 128.70, 122.24, 114.20, 55.84
3d	8.18(d, J =15.8 Hz, 1H, β -H), 8.03(s, 1H, Ph-H), 8.01(d, J =1.3 Hz, 1H, α -H), 7.76–7.74(m, 1H, Ph-H), 7.59(t, J =7.4 Hz, 1H, Ph-H), 7.50(dd, J =9.7, 5.7 Hz, 3H, Ph-H), 7.47(s, 1H, Ph-H), 7.45–7.42(m, 1H, Ph-H), 7.34–7.30(m, 2H, Ph-H)	190.32, 140.51, 137.81, 135.38, 133.14, 132.84, 131.08, 130.19, 128.56, 128.52, 127.68, 126.99, 124.69
3e	8.02(s, 1H, Ph-H), 8.01(d, J =1.3 Hz, 1H, Ph-H), 7.74(d, J =15.7 Hz, 1H, β -H), 7.60(t, J =7.4 Hz, 1H, Ph-H), 7.55(d, J =8.5 Hz, 2H, α -H), 7.53(d, J =6.2 Hz, 1H, Ph-H), 7.51(d, J =2.1 Hz, 3H, Ph-H), 7.50(s, 1H, Ph-H)	189.86, 142.99, 137.63, 133.43, 132.58, 131.84, 129.42, 128.31, 128.13, 124.43, 122.19
3f	8.05(s, 1H, Ph-H), 8.04(d, J =1.4 Hz, 1H, Ph-H), 7.91(d, J =15.9 Hz, 1H, β -H), 7.73(d, J =15.9 Hz, 1H, α -H), 7.59(t, J =7.4 Hz, 1H, Ph-H), 7.51(t, J =7.6 Hz, 2H, Ph-H), 7.23–7.21(m, 1H, Ph-H), 7.03(d, J =7.4 Hz, 1H, Ph-H), 7.01–6.97(m, 1H, Ph-H), 2.50(s, 3H, -CH ₃ -H)	190.77, 141.01, 140.93, 138.19, 136.00, 133.04, 130.66, 130.59, 128.69, 127.30, 127.21, 126.56, 122.25, 114.08, 113.93, 20.55
3g	8.10(d, J =15.8 Hz, 1H, β -H), 8.02–8.00(m, 2H, Ph-H), 7.68(d, J =8.5 Hz, 1H, α -H), 7.60(t, J =7.4 Hz, 1H, Ph-H), 7.52(d, J =7.9 Hz, 2H, Ph-H), 7.49(d, J =6.9 Hz, 1H, Ph-H), 7.46(d, J =2.4 Hz, 2H, Ph-H), 7.30(dd, J =8.4, 2.0 Hz, 1H, Ph-H)	190.24, 139.46, 137.90, 136.60, 136.20, 133.21, 131.98, 130.27, 128.84, 128.74, 128.63, 127.69, 125.15
3h	7.95(s, 1H, Ph-H), 7.94(s, 1H, Ph-H), 7.81(d, J =15.7 Hz, 1H, β -H), 7.65(dd, J =7.2, 2.2 Hz, 2H, Ph-H), 7.54(d, J =15.7 Hz, 1H, α -H), 7.42(d, J =2.1 Hz, 1H, Ph-H), 7.41(d, J =1.8 Hz, 1H, Ph-H), 7.31(d, J =8.0 Hz, 2H, Ph-H), 2.44(s, 3H, -CH ₃ -H)	190.15, 144.51, 135.76, 135.12, 130.54, 129.46, 129.06, 128.78, 128.53, 122.22, 21.80
3i	7.95(s, 1H, Ph-H), 7.93(s, 1H, Ph-H), 7.79(d, J =15.7 Hz, 1H, β -H), 7.55(s, 1H, Ph-H), 7.54(s, 1H, Ph-H), 7.50(d, J =15.7 Hz, 1H, α -H), 7.30(d, J =7.9 Hz, 2H, Ph-H), 7.22(d, J =7.9 Hz, 2H, Ph-H), 2.44(s, 3H, -CH ₃ -H), 2.39(s, 3H, -CH ₃ -H)	190.44, 144.81, 143.82, 141.26, 136.10, 132.60, 130.01, 129.63, 128.95, 128.76, 121.43, 21.99, 21.85

(Continued)

Compd.	¹ H NMR (600 MHz, CDCl ₃), δ	¹³ C NMR (150 MHz, CDCl ₃), δ
3j	8.12(d, <i>J</i> =15.9 Hz, 1H, β-H), 7.94(s, 1H, Ph-H), 7.93(s, 1H, Ph-H), 7.63(dd, <i>J</i> =13.2, 8.5 Hz, 2H, Ph-H, α-H), 7.37(t, <i>J</i> =8.6 Hz, 1H, Ph-H), 7.30(d, <i>J</i> =8.0 Hz, 2H, Ph-H), 6.99(t, <i>J</i> =7.5 Hz, 1H, Ph-H), 6.94(d, <i>J</i> =8.3 Hz, 1H, Ph-H), 3.91(s, 3H, -OCH ₃ -H), 2.43(s, 3H, -CH ₃ -H)	190.76, 158.89, 143.45, 140.07, 136.06, 131.75, 129.37, 129.29, 128.81, 124.18, 123.04, 120.85, 111.35, 77.37, 77.16, 76.95, 55.66, 21.79
3k	8.17(d, <i>J</i> =15.8 Hz, 1H, β-H), 7.93(d, <i>J</i> =8.2 Hz, 2H, Ph-H), 7.74(dd, <i>J</i> =7.0, 2.4 Hz, 1H, Ph-H), 7.48(d, <i>J</i> =15.7 Hz, 1H, α-H), 7.43(dd, <i>J</i> =6.8, 2.4 Hz, 1H, Ph-H), 7.31(d, <i>J</i> =4.2 Hz, 1H, Ph-H), 7.29(s, 2H, Ph-H), 2.43(s, 3H, -CH ₃ -H)	190.25, 144.21, 140.53, 135.80, 135.72, 133.73, 131.43, 130.64, 129.75, 129.15, 128.14, 127.44, 125.21, 22.07
3l	8.16(d, <i>J</i> =15.7 Hz, 1H, β-H), 8.03(d, <i>J</i> =8.9 Hz, 2H, Ph-H), 7.76–7.73(m, 1H, Ph-H), 7.49(d, <i>J</i> =15.7 Hz, 1H, α-H), 7.43(d, <i>J</i> =7.0 Hz, 1H, Ph-H), 7.31(d, <i>J</i> =4.3 Hz, 2H, Ph-H), 6.98(d, <i>J</i> =8.9 Hz, 2H), 3.89(s, 3H, -OCH ₃ -H)	189.60, 164.52, 140.75, 136.35, 134.45, 131.81, 131.25, 128.74, 128.01, 125.71, 114.87, 56.48

74% 产率获得 **3a** (表 3, Entry 1)。当浓硫酸用量增加到 20 equiv. 时, 产率升高 (表 3, Entry 2), 进一步增加浓硫酸用量, 产率降低 (表 3, Entry 3)。增加 PPA 用量对产率没有明显影响 (表 3, Entry 6), 但是 PPA 用量减小到 5 equiv. 时, 产率达到 96% (表 3, Entry 5), 进一步减少 PPA 添加量产率也随之降低 (表 3, Entry 4)。因此, 确定 PPA 和浓硫酸最佳用量分别为 5 equiv. 和 20 equiv.。之后考察了反应温度和反应时间对产率的影响, 改变反应温度和反应时间均使产率降低 (表 3, Entries 7~10)。当不添加 PPA 时无化合物 **3a** 生成 (表 3, Entry 11), 当不添加浓硫酸时以 8% 的产率获得化合物 **3a** (表 3, Entry 12)。由此可见 PPA 是催化苯乙酮和苯甲醛缩合生成查耳酮的必不可少的催化剂。然后, 用浓盐酸 (HCl)、磷酸 (H₃PO₄)、硝酸 (HNO₃) 或醋酸 (AcOH) 替换浓硫酸, 考察不同种类的酸与 PPA 共同催化的效果。替换酸后, 产率极低 (表 3, Entries 14~16), 甚至得不到目标产物 (表 3, Entry 13)。当使用乙醇 (EtOH)、四氢呋喃 (THF)、二甲基亚砜 (DMSO) 或 *N,N*-二甲基甲酰胺 (DMF) 作溶剂时, 均未获得目标产物 (表 3, Entries 17~20)。基于以上实验结果, 确定 PPA 5 equiv.、浓硫酸 20 equiv., 在二氧六环溶剂中, 于 90 °C 反应 2 h 为最优反应条件。

3.2 底物的拓展

为了探索该方法的适用范围, 在最优反应条件下, 以不同取代的苯乙酮和不同取代的苯甲醛为底物, 考察了不同取代基对反应产率的影响, 结果见表 1。当苯甲醛和/或苯乙酮取代基为供电子基甲基或甲氧基或无取代时 (**3a**~**3c**, **3h**~**3j**), 以高

Table 3 Optimization of the reaction conditions

Entry	Additive1 (equiv.)	Additive2 (equiv.)	Solvent	<i>T</i> /°C	<i>t</i> /h	Yield/%
1	PPA (10)	H ₂ SO ₄ (15)	1,4-dioxane	90	2	74
2	PPA (10)	H ₂ SO ₄ (20)	1,4-dioxane	90	2	86
3	PPA (10)	H ₂ SO ₄ (25)	1,4-dioxane	90	2	62
4	PPA (2.5)	H ₂ SO ₄ (20)	1,4-dioxane	90	2	72
5	PPA (5)	H ₂ SO ₄ (20)	1,4-dioxane	90	2	96
6	PPA (20)	H ₂ SO ₄ (20)	1,4-dioxane	90	2	87
7	PPA (5)	H ₂ SO ₄ (20)	1,4-dioxane	80	2	83
8	PPA (5)	H ₂ SO ₄ (20)	1,4-dioxane	100	2	85
9	PPA (5)	H ₂ SO ₄ (20)	1,4-dioxane	90	1	76
10	PPA (5)	H ₂ SO ₄ (20)	1,4-dioxane	90	3	80
11	PPA (0)	H ₂ SO ₄ (20)	1,4-dioxane	90	2	/
12	PPA (5)	H ₂ SO ₄ (0)	1,4-dioxane	90	2	8
13	PPA (5)	HCl (20)	1,4-dioxane	90	2	/
14	PPA (5)	H ₃ PO ₄ (20)	1,4-dioxane	90	2	8
15	PPA (5)	HNO ₃ (20)	1,4-dioxane	90	2	5
16	PPA (5)	AcOH (20)	1,4-dioxane	90	2	6
17	PPA (5)	H ₂ SO ₄ (20)	EtOH	90	2	/
18	PPA (5)	H ₂ SO ₄ (20)	THF	90	2	/
19	PPA (5)	H ₂ SO ₄ (20)	DMSO	90	2	/
20	PPA (5)	H ₂ SO ₄ (20)	DMF	90	2	/

THF: Tetrahydrofuran; DMSO: Dimethyl sulfoxide; DMF: *N,N*-dimethylformamide. Conditions were optimized with acetophenone (**1a**, 1.0 mmol) and benzaldehyde (**2a**, 0.8 mmol) as the substrate, and the solvent was 3.5 mL.

收率 (90%~96%) 得到目标化合物。当反应底物苯乙酮上有吸电子基溴时, 没有获得目标产物 (**3m**)。当苯甲醛上有吸电子基氟、氯或溴取代时产率降低 (**3d**~**3f**, **3k** 和 **3l**), 特别是当苯甲醛被两个吸电子基取代时产率进一步降低 (**3g**)。上述结果表明, 底物上有吸电子基取代时不利于该条件下缩

合生成查耳酮。

4 结 论

1,4-二氧六环为反应溶剂,于 90 °C 氮气保护下反应 2 h,PPA/H₂SO₄ 体系催化苯乙酮和苯甲醛缩合合成查耳酮,以高收率得到了 12 个查耳酮类化合物。该方法操作简单、反应条件温和、产率高且所使用催化剂成本低。该条件下合成的查耳酮类化合物均为 *E* 型,化合物 **3a~3l** 乙烯基质子 α -H (2-H) 和 β -H (3-H) 分别在 δ 7.4 和 δ 7.7 左右产生两组双重峰,偶合常数大于 15 Hz,与文献[18]报道的 *E* 型查耳酮 α -H、 β -H 化学位移一致。

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